

Survey of Aflatoxin M₁ Contamination of Dairy Products in Taiwan

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ABSTRACT

Owing to their nutritional value, dairy products were popular in Taiwan. Aflatoxin M₁(AFM₁) was the metabolite of potential carcinogen aflatoxin B₁ and found in dairy products. An analytical method using immunoaffinity column for extraction and HPLC for quantification was developed for AFM₁. The detection limits for fresh milk (pasteurized milk), milk powder and drinking yogurt (0.002, 0.02 and 0.005 ppb, respectively) were 10 times lower than that in the former survey. The recoveries of AFM₁ from fresh milk were 83.3 ± 2.9 and $89.5 \pm 2.9\%$ at 0.5 and 0.05 ppb spiked levels, respectively. Spiked 5 and 0.5 ppb AFM₁ in milk powder, the recoveries were 86.0 ± 1.9 and $88.7 \pm 1.9\%$, respectively. The recoveries of AFM₁ from drinking yogurt were 99.9 ± 1.4 and $94.8 \pm 3.2\%$ at 0.5 and 0.05 ppb spiked levels, respectively. It was the first time AFM₁ was tested in drinking yogurt in Taiwan in this survey, and the performance for detecting AFM₁ in drinking yogurt was also evaluated through attending an international proficiency test. Our laboratory got a satisfactory result. In order to survey AFM₁ contents in dairy products, 44 samples of fresh milk, 45 samples of milk powder and 24 samples of drinking yogurt were collected from supermarkets, convenience stores and drug stores located in 23 counties of Taiwan from June to August, 2002. The results showed that AFM₁ was detected in 40 samples of fresh milk at 0.002–0.083 ppb level. AFM₁ was not found in all milk powders. AFM₁ was detected in 3 samples of drinking yogurt, at the level of 0.007, 0.009 and 0.044 ppb. According to the food sanitary standard regulation in Taiwan, the action levels of AFM₁ were 0.5 ppb, 5 ppb and not-detectable for fresh milk, milk powder and infant formula products, respectively. The 113 samples collected in this survey all met the regulation requirements.

Key word: aflatoxin M₁, dairy product, immunoaffinity column, high performance liquid chromatography

INTRODUCTION

Aflatoxins, a group of several toxic secondary fungal metabolites produced by some *Aspergillus* spp., are found in a wide variety of foods and feeds around the world. Aflatoxin M₁ (AFM₁), a hydroxylated metabolite of the potential carcinogen Aflatoxin B₁ (AFB₁) (Figure 1), occurs in milk from lactating animals consuming feed contaminated with AFB₁⁽¹⁾. About 0.3–6.2% of AFB₁ in animal feed is transformed to AFM₁ in milk and a linear relationship has been found between intake of AFB₁ in contaminated feed and the AFM₁ content of milk in cows⁽²⁾. Following the withdrawal of the contaminated feed, AFM₁ levels in milk decrease to below the limit of detection within 72 hr⁽³⁾.

The potential hazardous human exposure to AFM₁ via consumption of milk and milk products has been demonstrated^(4,5,6,7). Both AFB₁ and AFM₁ can cause DNA

damage, gene mutation, chromosomal anomalies and cell transformation in mammalian cells *in vitro*, in insects, lower eukaryotes and bacteria⁽⁸⁾. However, AFM₁ is less carcinogenic and genotoxic than AFB₁⁽²⁾.

To protect consumers, particularly children, from contaminated dairy products, several countries have established legislation to regulate the levels of AFB₁ in feeds and AFM₁ in milk (Table 1). The Food and Drug Administration (FDA) of US has established an action level of 0.50 ppb in whole, low fat and skim milk⁽⁹⁾, whereas the EU has set a maximum admissible level of 0.05 ppb in raw milk, heat-treated milk, and milk for the manufacture of milk based products⁽¹⁰⁾. In Taiwan, the action levels in fresh milk, milk powder and infant formula dairy products are 0.5 ppb, 5 ppb and not-

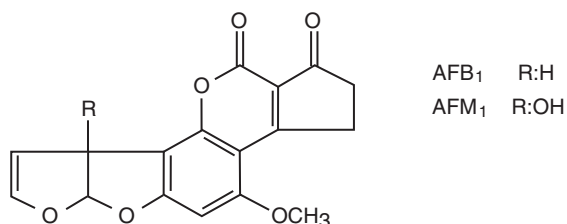


Figure 1. Chemical structures of AFB₁ and AFM₁.

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Table 1. Regulatory limits for AFM₁ in various countries

| Country | Milk (ppb) | Infant formula (ppb) |
|-------------|------------------|----------------------|
| Argentina | 0.5 ^a | 0.1 |
| Austria | 0.05 | 0.01 |
| Brazil | 0.5 | 0.01 |
| France | 0.2 | |
| Germany | 0.05 | 0.01 |
| Italy | 0.05 | 0.05 |
| Netherlands | 0.05 | 0.05 |
| Switzerland | 0.05 | 0.01 ^b |
| US | 0.50 | |
| EU | 0.05 | |
| Taiwan | 0.5 | Not-detectable |

^aSum of B₁, B₂, G₁, G₂ and M₁.

^bSum of B₁ and M₁.

detectable, respectively^(11,12).

Monitoring surveys are frequently conducted in several countries all over the world to determine the levels of AFM₁ in milk and milk products. In Taiwan, 161 raw milk and milk powder had been examined according to the AOAC method in 1986, and no AFM₁ contamination in these samples was found⁽¹³⁾. Using a silica-gel column to extract and clean up all 110 dairy samples, 44 of 50 (88%) fresh milk, 15 of 25 (60%) milk powder, 6 of 10 (60%) condensed milk and 0 of 25 cheese samples were detected the contamination of AFM₁⁽¹⁴⁾. Fu determined the AFM₁ in milk and milk powder using an immunoaffinity column and fluorescence measurement. AFM₁ was detected in 3 of 25 (12%) fresh milk and 0 of 25 milk powder samples⁽¹⁵⁾. However, no survey has referred to the occurrence of AFM₁ in drinking yogurt. The objectives of this work were to establish a more sensitive method to determine the AFM₁ in dairy products (especially in drinking yogurt) and to understand the contamination of AFM₁ in fresh milk, milk powder and drinking yogurt in Taiwan.

MATERIALS AND METHODS

I. Materials

(I) Sample collection

The feed in Taiwan is much easier to be contaminated with AFB₁ due to the warm and humid weather in summer; therefore, this survey was conducted during June ~ August. The samples were collected from supermarkets, convenience stores and drug stores of 23 counties in Taiwan. Forty-four samples were fresh milk, 45 samples were milk powder (of which 21 were infant formula) and 24 samples were drinking yogurt. The minimum sample size was 500 mL or g. The samples were conserved in a refrigerator during the transfer to the laboratory. In the laboratory, fresh milk and drinking yogurt samples were stored in a refrigerator ($5 \pm 2^\circ\text{C}$), and milk powder samples were stored at a temperature below 25°C . All samples were analyzed before the expiration date.

(II) Reagents

AFM₁ standard was purchased from Supelco (Bellefonte, PA., U.S.A.). The immunoaffinity columns AflaM₁TM were purchased from Vicam (Watertown, MA., U.S.A.). Sodium hydroxide was reagent grade. Acetonitrile and methanol used for the liquid chromatographic mobile phases were of LC grade. Distilled, deionized water was used throughout the procedure.

II. Methods

(I) Samples preparation

The procedures were modified from Dragacci *et al.*⁽¹⁶⁾

and Tuinstra *et al.*⁽¹⁷⁾ Fresh milk was warmed before analysis to ca 37°C in a water bath, and then stirred gently with a magnetic stirrer to disperse the fat layer. Ten grams of milk powder (including infant formula) were dissolved with 50 mL of warm water, and mixed with stirring until a homogeneous mixture was obtained. The solution of milk powder was transferred to a 100-mL volumetric flask and diluted to the mark. Liquid milk (fresh milk and the solution of milk powder) was centrifuged for 10 min at $2000\times g$ to separate the fat, and the upper fat layer was discarded. The samples were filtered through one or more paper filters, and at least 50 mL of the filtrate was collected. Twenty-five grams of drinking yogurt was warmed as fresh milk and the pH was adjusted to 6.5 with 0.1N NaOH before passing samples through the clean-up column.

(II) Immunoaffinity clean up

Fifty mL of liquid milk and pH-adjusted drinking yogurt were passed at about 1-2 mL/min through an AflaM₁TM immunoaffinity column fitted with a 60-mL syringe reservoir. Twenty mL of water was used to wash the loaded immunoaffinity column at a steady flow rate. The column was blown to dryness with a stream of nitrogen, and AFM₁ was eluted with 4 mL of acetonitrile. The acetonitrile eluate was evaporated to dryness under a gentle stream of nitrogen, and the residue was redissolved by vortexing with 2 mL of mobile phase, which was then filtered through a $0.45 \mu\text{m}$ microfilter for HPLC analysis.

(III) HPLC analysis

The LC system consisted of a Shimadzu (Kyoto, Japan) LC-10AT_{VP} pump, a RF-10A_{XL} fluorescence detector, and a Waters 717_{plus} autosampler (Waters, MA., U.S.A.), all under the control of the Shimadzu SCL-10A_{VP} system. Data acquisition was performed on an SISC program. The column ($150 \times 4.6 \text{ mm}$, Cosmosil 5C18-AR, $5 \mu\text{m}$, Nacalai, Japan) was maintained at 30°C . Injection volume was 200 μL and the detector wavelength settings were 365 nm (excitation) and 435 nm (emission). The mobile phase, water/acetonitrile/methanol (68/24/8, v/v/v), was pumped at a constant flow rate of 1.0 mL/min.

(IV) Calibration

AFM₁ standard was diluted with mobile phase to prepare a series of working solutions containing 0.002-10 ng AFM₁/200 μL . A calibration curve was constructed by plotting the peak area for each standard against the mass of AFM₁ injected. Slope and intercept data of the calibration curve were used to compute the quantity of the analyte in sample extracts.

(V) Validation

Analytical methods for AFM₁ determination from

three kinds of dairy products were validated in-house. Two each of AFM₁-free fresh milk and drinking yogurt samples were spiked with 0.05 and 0.5 ppb AFM₁ standards. Two AFM₁-free milk powders were spiked with 0.5 and 5 ppb AFM₁ standards, then dissolved with warm water. Then the following procedures were according to those described above. The method detection limit (MDL) of the chromatographic procedure was measured according to the Standard Operation Procedure for the MDL determination of Bureau of Food and Drug Analysis⁽²⁹⁾ as follows:

AFM₁ standards of serial concentrations were injected into and analyzed by the LC system. A curve was constructed by plotting the peak area for each standard against the mass of AFM₁ injected. The concentration where the slope of the standard curve changed markedly was defined as IDL (instrument detection limit). To obtain the MDL, an AFM₁ standard at a concentration equal to 1~5 times IDL was spiked into AFM₁-free samples, followed by the analytical procedures described above. Standard deviation (Sa) was calculated from 7 replicates. An AFM₁ standard at the concentration equal to 3 times Sa was spiked into AFM₁-free samples, followed by the analytical procedures described above. Standard deviation (S) was calculated from 7 replicates. The bigger one of Sa² and S² was chosen as the numerator. If the ratio was smaller than 3.05, the pooled standard deviation (S_{pool}) and MDL were calculated using the following regulations :

$$S_{pool} = [(6S^2 + 6Sa^2) / 12]^{1/2}$$

$$MDL = 2.681 \times S_{pool}$$

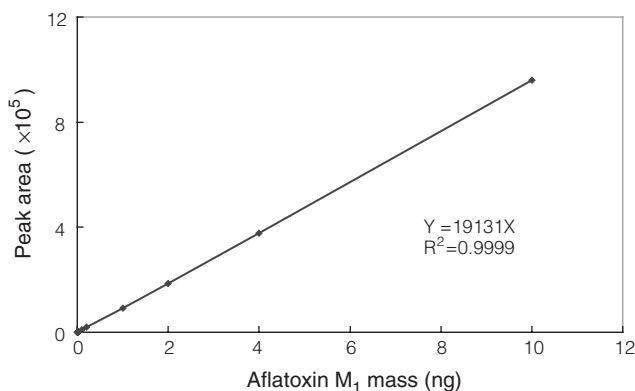


Figure 2. Calibration curve of AFM₁ analyzed by HPLC.

Table 2. Recoveries of AFM₁ in dairy products

| Dairy product | Spiked levels (ppb) | Recovery ^a (%) | CV ^b (%) |
|-----------------|---------------------|---------------------------|---------------------|
| Fresh milk | 0.05 | 89.5 ± 2.9 | 3.2 |
| | 0.5 | 83.3 ± 2.9 | 3.5 |
| Drinking yogurt | 0.05 | 94.8 ± 3.2 | 3.4 |
| | 0.5 | 99.9 ± 1.4 | 1.4 |
| Milk powder | 0.5 | 88.7 ± 1.9 | 2.1 |
| | 5 | 86.0 ± 1.9 | 2.2 |

^aAverage of triplicate analysis ± standard deviation.

^bCV: coefficient of variation.

(VI) Quantification

AFM₁ mass concentration of the test sample was calculated using the following equation:

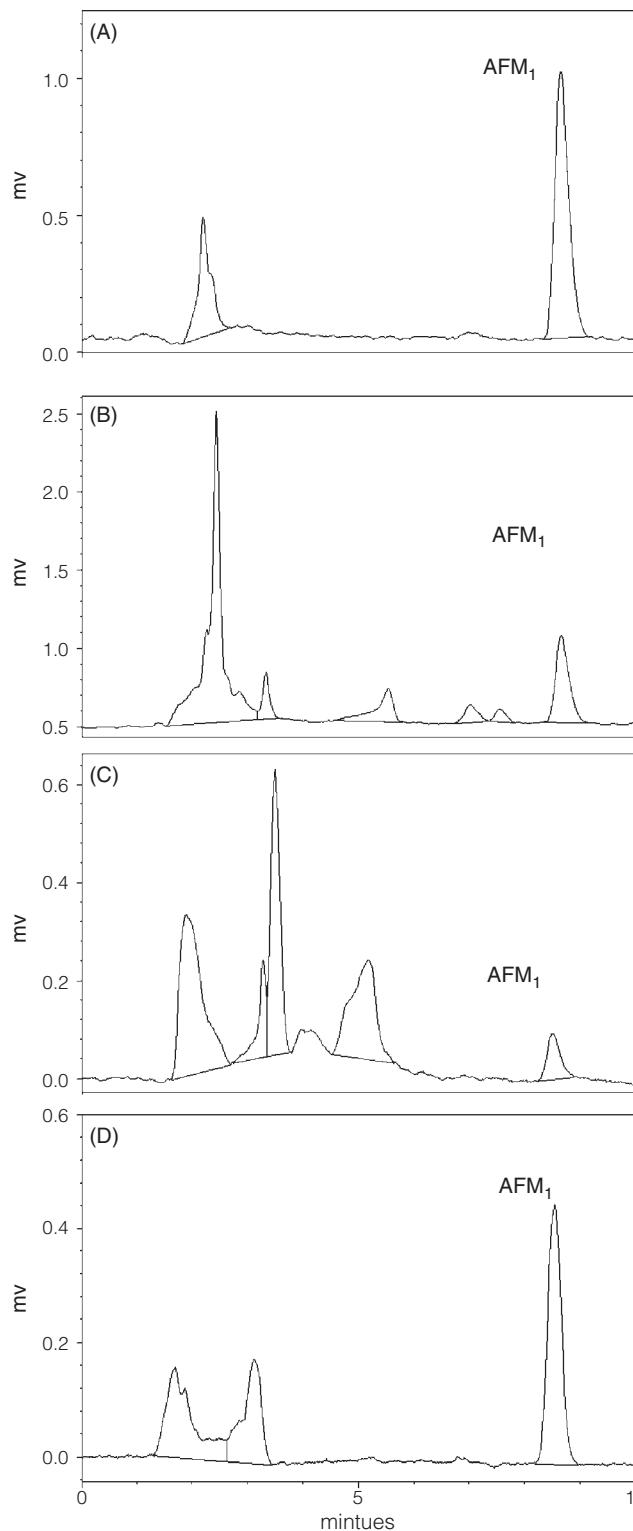


Figure 3. HPLC chromatograms of (A) AFM₁ standard, (B) AFM₁-spiked fresh milk, (C) AFM₁-spiked milk powder, (D) AFM₁-spiked drinking yogurt. HPLC conditions: Cosmosil 5C18-AR; mobile phase, water/acetonitrile/methanol (68/24/8, v/v/v) flow rate, 1.0 mL/min.

$$W_m = W_a \times (V_f/V_i) \times (1/V_s)$$

Where W_m = the numerical value of AFM₁ mass concentration in the test sample in ppb (ng/g or ng/mL); W_a = the numerical value of the amount of AFM₁ corresponding to the area of AFM₁ peak of the sample extract (ng); V_f = the numerical value of the final volume of redissolved eluate (μL); V_i = the numerical value of the final volume of injected eluate (μL); V_s = the numerical value of volume or mass of prepared test portion passing through the column (mL or g).

Express the results to 3 significant figures.

(VII) Proficiency test

Our Laboratory participated in the “Analysis of Aflatoxin M₁ in yogurt” (Food Analysis Performance Test Assessment Scheme, Series 4, Aflatoxins Analysis, Round 4) proficiency test held by the Central Science Laboratory (Sand Hutton, York, U.K.) in February, 2002. The sample was analyzed as described above.

RESULTS

I. Analytical Method Performance

(I) Calibration

Within the calibration range of 0.002-10 μg/200 μL of AFM₁, the HPLC responses (peak area) positively regressed with injected AFM₁ mass and gave a correlation coefficient of $R^2 = 0.9999$, as shown in Figure 2.

(II) Validation

The MDLs of fresh milk, milk powder and drinking yogurt were 0.002, 0.02 and 0.005 ppb, respectively. Specificity of the AFM₁ peaks were clearly shown in HPLC chromatograms of Figure 3, which showed the absence of interfering signals at the AFM₁ retention area for fresh milk, milk powder and drinking yogurt. The recoveries of AFM₁ in fresh milk, milk powder and drinking yogurt were 83.3-89.5, 86.0-88.7, and 94.8-99.9%, respectively, as shown in Table 2.

(III) Proficiency test

Thirty-four of 43 participants delivered their detection results and 25 participants received the “satisfied result” ($|z| < 2$). The z-scores of all 34 participants were shown in Figure 4. Our laboratory number was 7, and the z-score was 0.7.

II. Survey Results (Table 3)

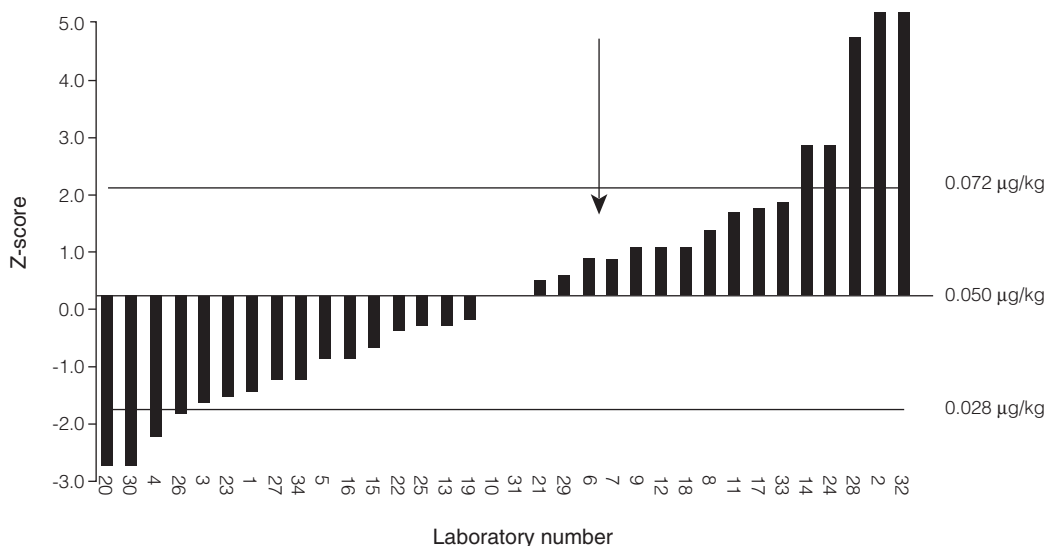


Figure 4. Z-score distribution of participants in the “Analysis of AFM₁ in yogurt” proficiency test. (The number of our laboratory was 7 as the arrow indicated)

Table 3. AFM₁ contents of dairy products

| Dairy product | No. of samples | No. of positive samples (%) | AFM ₁ contents (ppb) | | |
|-----------------|----------------|-----------------------------|---------------------------------|-----------|----------|
| | | | 0.002~0.01 | 0.01~0.05 | 0.05~0.1 |
| Fresh milk | 44 | 40 (90.9) | 10 | 29 | 1 |
| Milk powder | 24 | 0 (0) | 0 | 0 | 0 |
| Infant formula | 21 | 0 (0) | 0 | 0 | 0 |
| Drinking yogurt | 24 | 3 (12.5) | 2 ^a | 1 | 0 |
| Total | 113 | 43 (38.1) | 12 | 30 | 1 |

^aBetween 0.007 and 0.01 ppb.

Table 4. Incidence and levels of AFM₁ contamination in fresh milk

| Country | Year | No. of samples | No. of positive samples (%) | AFM ₁ (ppb) |
|-------------------------------------|-----------|----------------|-----------------------------|---------------------------|
| Brazil ⁽¹⁹⁾ ^a | 1992 | 52 | 4 (7.7) | 0.073 ~ 0.370 |
| Italy ⁽²⁰⁾ | 1995 | 159 | 136 (85.5) | < 0.001 ~ 0.11 |
| Italy ⁽²¹⁾ | 1996 | 161 | 125 (77.6) | < 0.001 ~ 0.024 |
| Tailand ⁽²²⁾ | 1995~1996 | 250 | 249 (99.6) | 47 more than 0.5 ppb |
| Taiwan ⁽¹⁵⁾ | 1996 | 25 | 3 (12.0) | 0.08 ~ 0.33 |
| Korea ⁽⁷⁾ | 1997 | 70 | 39 (55.7) | 0.015 ~ 0.052 |
| Kuwait ⁽²³⁾ | 1998 | 30 | 13 (43.3) | 0.01 ~ 0.21 |
| Spain ⁽²⁵⁾ | 1998 | 47 | 14 (29.8) | 0.02 ~ 0.1 |
| Portugal ⁽²⁶⁾ | 1999 | 101 | 85 (84.2) | 2 more than 0.05 ppb (2%) |
| Greece ⁽²⁴⁾ | 1999~2001 | 136 | 113 (83.0) | 0.005 ~ 0.05 |
| Brazil ⁽²⁷⁾ | 1999~2000 | 139 | 29 (20.9) | 0.05 ~ 0.24 |
| Taiwan ^b | 2002 | 44 | 40 (90.9) | 0.002 ~ 0.083 |

^aReference.

^bData of this survey.

Table 5. Incidence and levels of AFM₁ contamination in milk powder

| Country | Year | No. of samples | No. of positive samples (%) | AFM ₁ (ppb) |
|------------------------------------|-----------|----------------|-----------------------------|------------------------|
| Italy ⁽²⁰⁾ ^a | 1995 | 97 | 81 (83.5) | < 0.001 ~ 0.10 |
| Italy ⁽²¹⁾ | 1996 | 92 | 50 (54.3) | < 0.001 ~ 0.08 |
| Tailand ⁽²²⁾ | 1995~1996 | 13 | 2 (15.4) | Not more than 0.5 ppb |
| Taiwan ⁽¹⁵⁾ | 1996 | 25 | 0 | 0 |
| Korea ⁽⁷⁾ | 1997 | 50 | 35 (70.0) | 0.032 ~ 0.342 |
| Brazil ⁽²⁸⁾ | 1997 | 300 | 33 (11.0) | 0.1 ~ 1.0 |
| Kuwait ⁽²³⁾ | 1998 | 19 | 0 | 0 |
| Taiwan ^b | 2002 | 45 | 0 | 0 |

^aReference.

^bData of this survey.

Table 6. Incidence and levels of AFM₁ contamination in yogurt

| Country | Year | No. of samples | No. of positive samples (%) | AFM ₁ (ppb) |
|------------------------------------|------|----------------|-----------------------------|------------------------|
| Italy ⁽²⁰⁾ ^a | 1995 | 114 | 91 (79.8) | < 0.001 ~ 0.50 |
| Italy ⁽²¹⁾ | 1996 | 120 | 73 (60.8) | < 0.001 ~ 0.032 |
| Korea ⁽⁷⁾ | 1997 | 60 | 31 (51.7) | 0.017 ~ 0.124 |
| Kuwait ⁽²³⁾ | 1998 | 5 | 1 (20.0) | 0.01 ~ 0.21 |
| Taiwan ^b | 2002 | 24 | 3 (12.5) | 0.007 ~ 0.044 |

^aReference.

^bData of this survey.

(I) Fresh milk

Among the 44 fresh milk samples, 3 samples were manufactured by small scale farms and sold only in local supermarkets. The remaining 41 samples were manufactured by larger scale food companies and sold to other counties. AFM₁ above the MDL of 0.002 ppb was present in 40 of 44 (90.9%) fresh milk samples, among which 10 were between 0.002 and 0.01 ppb, 29 were between 0.01 and 0.05 ppb, 1 was between 0.05 and 0.1 ppb (0.083 ppb).

(II) Milk powders and infant formula

The milk powders and infant formula were all goods imported. No AFM₁ was detected in the 24 milk powders and 21 infant formula according to the MDL of 0.02 ppb.

(III) Drinking yogurt

Two of 24 samples of drinking yogurt were manufac-

ured by small scale food processing companies and sold only in local supermarkets. The remaining 22 samples were manufactured by larger scale food companies and sold to other counties. The incidence of AFM₁ in drinking yogurt (12.5%) was lower than that in fresh milk. Three of 24 drinking yogurt samples were contaminated by AFM₁ at the concentrations of 0.007, 0.009 and 0.044 ppb.

DISCUSSION

The retention time of AFM₁ in the HPLC chromatograms was shorter in this survey (8~9 min) than that in the former survey (13~14 min)⁽¹⁵⁾. The MDL was 10 times lower.

Fu⁽¹⁵⁾ surveyed the AFM₁ content in fresh milk, and found that 3 of 25 (12%) samples were positive. The column Fu used (AFLAPREP M, Vicam) was different from what we used in this survey, and the MDL was 0.05 ppb, which was much higher than those in this survey.

Though the principles of these two surveys were similar, the sensitivity of the HPLC system in this survey was elevated and the MDL was lowered to 0.002 ppb. According to the MDL of 0.05 ppb, only 1 of 44 (2.3%) fresh milk was found to be positive in this survey. Hence, the AFM₁ contamination status in fresh milk was not serious. According to Visconti *et al.*⁽¹⁸⁾, most of such results should be attributed to the poor sensitivity of the analytical methods used. Today, the increased efficiency of the extraction procedures and the increased accuracy of the analytical methods and better equipments resulted in much lower limits of detection, thereby a significant increase in the percentage of positive samples.

In comparison, the incidence of AFM₁ contamination in fresh milk in recent studies ranged from 7.7 to 99.6%, with means of AFM₁ contents from 0.01 to 0.5 ppb, as shown in Table 4^(7,15,19,20,21,22,23,24,25,26,27). In this survey, the incidence of AFM₁ contamination in fresh milk was very high, 90.9% of the samples were positive. However, Table 3 showed that AFM₁ was not detected in 9.1% of the fresh milk sample, detected at low levels (0.002 to 0.05 µg/L) in 88% of the samples and none of these positive samples were above the action level of 0.5 ppb. This supports the view that a high incidence of low-level contamination and infrequent high contamination levels in commercial milk could be caused by only a few contaminated samples entering the bulk milk supply⁽²⁰⁾.

The incidence of AFM₁ in milk powder in recent studies ranged from 0 to 84%, with AFM₁ mean contents from 0 to 1.0 ppb, as shown in Table 5^(7,15,20,21,22,23). The 45 milk powder samples (including 21 infant formula) collected in this survey were all imported from America, Japan, New Zealand and the Netherlands. The same as the result of a former survey held in 1996⁽¹⁵⁾, no AFM₁ was detected in the milk powder samples. Though there were trace levels of AFM₁ contamination found in the milk powder samples in the recent survey, none of these positive samples was detected at levels above the action level of 5.0 ppb, and it seems that the milk powders exported to Taiwan were free of AFM₁. Since Taiwan became a member of WTO, there will be more and more milk powders imported from different countries to Taiwan. Therefore, related surveys should be continued in the future.

Owing to its nutritional value, yogurt, especially drinking yogurt, has become more and more popular in Taiwan. However, the content of AFM₁ is not reduced in the fermentation process if AFM₁-contaminated milk is used. The concentration of AFM₁ could even be increased due to the condensation process⁽²⁰⁾. The incidence of AFM₁ in yogurt in recent studies ranged from 20 to 80%, with AFM₁ mean contents from 0 to 0.50 ppb, as shown in Table 6^(7,20,21,22,23). In this survey, it was the first time AFM₁ was tested in drinking yogurt in Taiwan. Though there was AFM₁ contamination in drinking yogurt, the content was low (0.007 to 0.044 ppb). Till now, the action level of AFM₁ in yogurt has not been set in Taiwan.

CONCLUSION

By combining immunoaffinity column for cleanup and LC with fluorescence for detection, the AFM₁ contents in dairy products could be measured with good recoveries and satisfactory MDL. Through participation in the "Analysis of Aflatoxin M₁ in yogurt" in 2002, our laboratory was among the 34 laboratories which received "satisfied result" in the proficiency test.

In this survey, it was revealed that trace levels of AFM₁ were detected in dairy products such as fresh milk and drinking yogurt in Taiwan. However, none of these positive samples was above the action level of 0.5 ppb.

ACKNOWLEDGMENTS

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