藥物食品分析第二卷 第二期

Characterization of ¹⁴C Terminal Residues in Rice Plants Treated with ¹⁴C Ring-Labeled Benthiocarb

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ABSTRACT

The terminal ¹⁴C residues in rice treated with ¹⁴C ring-labeled benthiocarb were characterized. Rice was treated at 5 1b/acre, then grown to maturity. Straw and grain were harvested and analyzed for ¹⁴C. The total ¹⁴C in the straw and the grain was 2.0 and 0.3 ppm, respectively, calculated as benthiocarb. No (less than 0.01 ppm) parent compound or metabolites containing the intact thiocarbamate moiety were found in either the straw or the grain. Approximately 0.25 ppm 4-chlorobenzoic acid (0.41 ppm ¹⁴C calculated as benthiocarb) was isolated from straw as bound, conjugated and/or free metabolites. Two other major ¹⁴C metabolites (0.33 ppm), both containing carboxylic acid and the 4-chlorobenzylthio moieties in the molecules, were partially identified. Approximately 0.02 ppm 4-chlorobenzyl methyl sulfone was found in the grain.

Key words: Thiocarbamate herbicide, Degradation products of benthiocarb

INTRODUCTION

Benthiocarb [S-4-chlorobenzyl N, N-diethyl-thiocarbamate] is a thiocarbamate with good herbicidal activity. This chemical has been used for the last two decades to control weeds in rice paddies. Benthiocarb is particularly effective against barnyard grass there. A few studies have been conducted on the metabolic and the environmental fates of this chemical. They include: animal metabolism⁽¹⁻³⁾; plant metabolism^(2, 4-5); microbial and soil metabolism⁽⁶⁻⁸⁾; photodegradation⁽⁸⁻⁹⁾; and degradation in water environment⁽¹⁰⁻¹¹⁾. The fate and Persistence of benthiocarb and its potentially toxic metabolite in mature rice plant have not yet been described.

The purpose of this study was to investi-

gate the residue and the degradation products of benthiocarb in mature rice. One week after seeding, rice was treated with ¹⁴C ring-labeled benthiocarb at 5 lb/acre, then grown to maturity (5 months). Straw and grain were harvested, and the ¹⁴C residues in the samples were characterized.

MATERIALS AND METHODS

I .Chemicals

The [phenyl-U- 14 C] benthiocarb was purchased from New England Nuclear (Boston, Massachusette). The 14 C ring-labeled benthiocarb which had a specific activity of 3.68 mCi/mM (31,788 dpm/ μ g) was used. All authentic compounds for cochromatography used in this study

were: N-ethyl-S-(4-chlorobenzyl)thiocarbamate, S -(4-chlorobenzyl)thiocarbamate, N,N-diethyl-S-(4-chloro-2-hydroxybenzyl)thiocarbamate, Nethyl-S-(4-chloro-2-hydroxybenzylthiocarbamate, N,N-diethyl-S-(4-chloro-3-hydroxybenzyl) ocarbamate, N,N-diethyl-S-(4-chlorobenzyl)-Smonoxythiocarbamate, N,N-diethyl-S-(4-chlorobenzyl)-S-dioxythiocarbamate, 4-chlorobenzyl alcohol, 2-hydroxyl-4-chlorobenzyl alcohol, 3-hydroxyl-4-chlorobenzyl alcohol, 4-chlorobenzoic acid, 2-hydroxy-4-chlorobenzoic acid, 3-hydroxy-4-chlorobenzoic acid, 4-chlorobenzyl mercaptan, 4-chlorobenzyl sulfonic acid, bis(4-chlorobenzyl) disulfide, bis(4-chlorobenzyl) monoxydisulfide, bis(4-chlorobenzyl) dioxydisulfide, 4-chlorobenzyl methyl sulfide, 4-chlorobenzyl methyl sulfoxide, 4-chlorobenzyl methyl sulfone, N-4-chlorobenzoylglycine, N-4-chlorobenzoylleucine, 4hydroxybenzoic acid, 3,4-dihydroxybenzoic acid, 3,4,5-trihydroxybenzoic acid, 4-hydroxycinnamic acid, caffeic acid, vanillic acid, ferulic acid and syringic acid. All these authentic compounds were supplied by the Organic Synthesis Division, Chevror Chemical Company, Richmond, California.

II. Planting and Treatment of Rice

A stock watering tank (6 \times 8 ft.) was used for growing the rice. The tank was placed outside, uncovered at the Delta Branch Experimental Station, Mississippi State University, Stoneville, Mississippi. Soil was placed in the tank to a depth of six inches. An overflow pipe was four inches above the soil level. Rice (Nato variety) was planted at a rate of 400 1b seed/acre. Treatment with formulated ¹⁴C ring-labeled benthiocarb at 5 1b/acre was carried out 1 week later. The formulated material was prepared by adding 1.161 g of ¹⁴C ring-labeled benthiocarb 117 mg Atloz emulsifying agents, 3409F/3404F (60/40) in ether. The ether was evaporated and 23 mg of xylene added to mixture. The formulated benthiocarb was diluted to 50 ml with water, and 25 ml of this solution was applied to the tank.

Flood water was added three weeks after

treatment when rice was four to six inches tall. The rice was grown under flooded conditions for 5 months after planting up to harvest.

III. Assay for the ¹⁴C

All extracts and regions scraped from thin-layer chromatoplates were counted in a Nuclear Chicago Isocap/300 liquid scintillation spectrometer. Aliquots (0.01-1 ml) of the various extracts or silica gel regions scraped from chromatoplate were dissolved in 10 ml of Scintisol (Isolab, Inc.) or 16 ml of a scintillation mixture containing 8 ml 2-methoxyethanol and 8 ml PPO (2,5-diphenyloxazole) in toluene (8 g/l). Solid samples ranging from 50 to 200 mg before and after extraction were combusted in duplicate with a Packard Model 306 sample oxidizer. The combusted samples were added to the scintillation mixture and counted in a liquid scintillation counter for determination of the ¹⁴C.

IV.Extraction of the ¹⁴C

Samples of straw or grain weighing 25 to 50 g were extracted three times with 200 ml portions of methanol-water 1:1 (v/v). The extractions were carried out in a Virtis Homogenizer for 20 minutes. The extraction flask was placed in a water bath at about 50°C. The residue remaining after methanol-water extraction was further extracted with aqueous 0.5 N sodium hydroxide. The extraction procedure is shown in Figure 1. The lignin preparation was carried out according to the procedure described by Browing⁽¹²⁾.

V.Fractionation and Separation of the ¹⁴C

The methanol was removed from the aqueous methanol extract with a rotary evaporator. The aqueous residue was acidified to pH 1 with 20% HCl, then saturated with (NH₄)₂SO₄. The ¹⁴C in this mixture was separated into five fractions; i.e. hexane, ether, ether-ethanol (2:1), aqueous residue and brown solid. Each of the

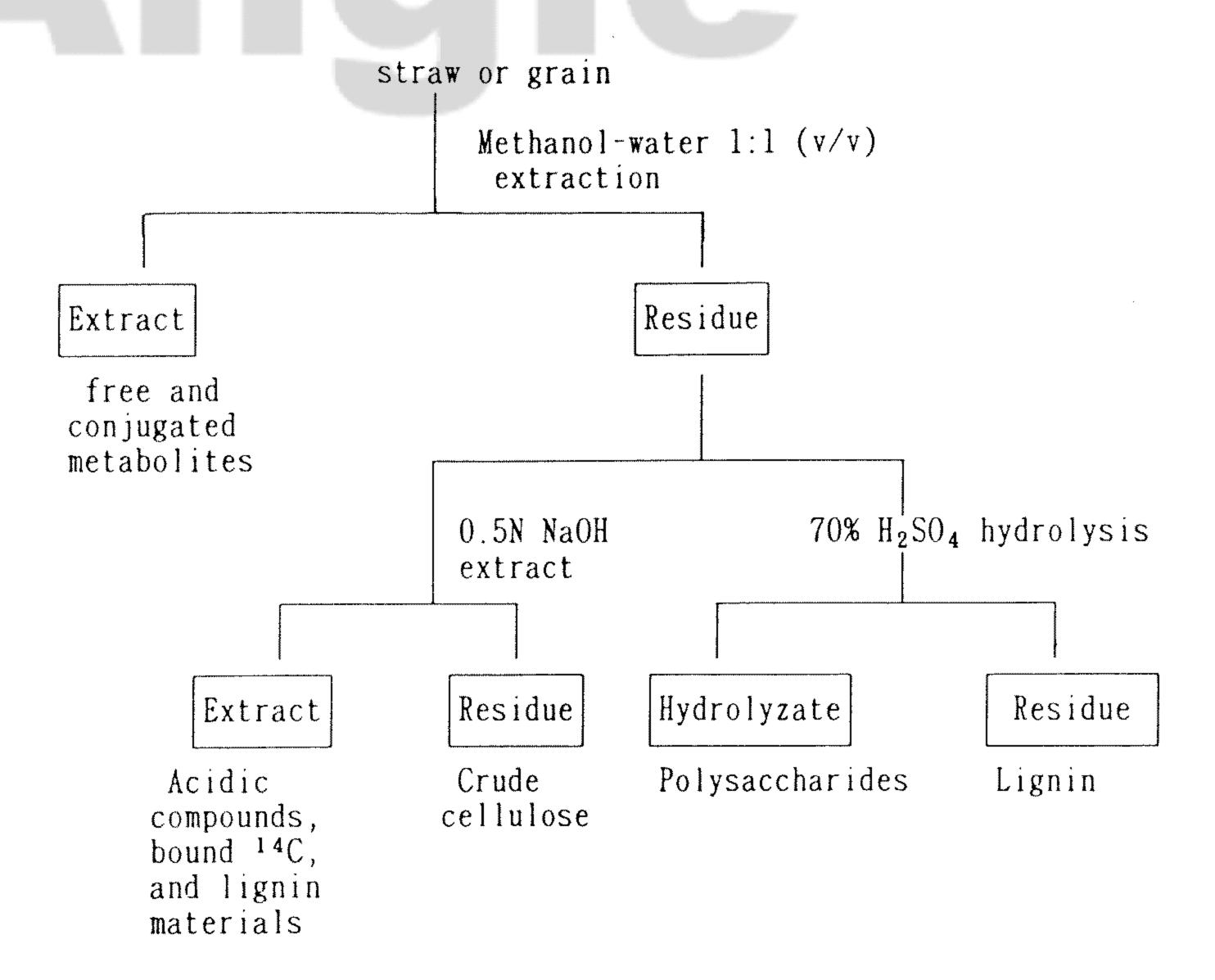


Figure 1. Extraction procedure for separation of ¹⁴C in straw and grain.

organo-extractable fractions was further separated into acid and neutral or phenolic fractions by extraction with 5% KHCO₃. The fractionation procedure is shown in Figure. 2.

VI.Isolation and Identification of the ¹⁴C

Thin-layer chromatography (TLC) utilized 20 × 20 cm silica gel F-254 precoated chromatoplates (E.M. Laboratories, Inc., Elmsfor, N.Y.) with 0.25 mm or 0.5 mm gel thickness for analysis. The solvent systems used for isolation of the ¹⁴C degradation products and for cochromatography with authentic compounds were as follows: (A) hexane only; (B) hexane-benzene (1:1); (C) chloroform-ether (4:1); (D) benzeneether (1:1); (E) benzene (saturated with formic acid)-ether (3:1); (F) benzene-methanol-formic acid (100:20:1). Rf values for the authentic compounds using one dimensional development are given in Table 1. Various combinations and sequences of developing solvents were used for adequate resolution of the metabolite. Radioautography with single coated blue sensitive X-ray

film (Eastman Kodak Co., Rochester, N.Y.) was used to locate radioactive derivatives on TLC chromatoplate.

VII.Enzyme Hydrolysis

Samples were incubated with β -glucosidase (1000 units/mg, 50 mg, Calbiochem, San Diego, CA) in acetate buffer, pH 4.6, at 34 °C overnight. The incubation mixture was acidified with HCl to pH 1 and extracted with ether. The ether extract was characterized by TLC analysis.

RESULTS

Extraction Study

Various extraction mediums were tested, and showed that methanol-water 1:1 (v/v) was the most effective solvent for the extraction of the ¹⁴C in the straw and grain. It is sufficient for extraction of both free and conjugated metabolites. Approximately 50% (1 ppm) of the ¹⁴C in the straw and 10% (0.03 ppm) in the grain

were extracted with aqueous methanol.

Nature of the ¹⁴C in the Straw (2 ppm)

About (50%) 1 ppm of the ¹⁴C was extracted with neutral aqueous methanol. About 0.7 ppm (35%) of the remaining ¹⁴C was further extracted with 0.5 N sodium hydroxide. About 0.2 ppm (10%) of the ¹⁴C was not extracted.

Characterization of the ¹⁴C in the Methanol-Water Extract from Straw (1.1 ppm)

To facilitate the identification of the ¹⁴C by TLC, the ¹⁴C in the extract was separated into several fractions and characterized. The ¹⁴C distribution is given in Table 2. Those fractions were:

1. Hexane Soluble ¹⁴C

Over 95% (0.08 ppm) of this ¹⁴C were acidic compounds extractable with 5% potassium bicarbonate solution. Most (0.08 ppm) of the ¹⁴C was identified as 4-chlorobenzoic acid

Table 1. Thin-layer chromatography Rf values for standard compounds

Compound	Rf	values	in	indicated	TLC	solvent	system
		A	В	С	D	E	F
4-chlorobenzoic acid		0.00	0.00	0.12	0.50	0.49	0.38
2-hydroxy-4-chlorobenzoic acid		0.00	0.00	0.05	0.13	0.52	0.24
3-hydroxy-4-chlorobenzoic acid		0.00	0.00	0.05	0.24	0.32	0.29
N-4-chlorobenzoylglycine		0.00	0.00	0.00	0.00	0.12	0.22
N-4-chlorobenzoylleucine		0.00	0.00	0.05	0.13	0.25	0.30
N,N-diethyl-S-(4-chlorobenzyl)		0.00	0.12	0.60	0.91	0.60	0.65
thiocarbamate							
N-ethyl-S-(4-chlorobenzyl) thiocarbamate		0.00	0.07	0.53	0.88	0.55	0.57
S-(4-chlorobenzyl)thiocarbamate		0.00	0.01	0.30	0.71	0.40	0.40
4-chlorobenzyl alcohol		0.00	0.04	0.30	0.63	0.35	0.40
2-hydroxy 4-chlorobenzyl alcohol		0.00	0.00	0.23	0.60	0.30	0.35
4-chlorobenzyl mercaptan		0.22	0.45	0.65	0.94	0.67	0.67
bis-4-chlorobenzyldisulfide		0.17	0.50	0.65	0.94	0.67	0.67
4-chlorobenzyl methyl sulfide		0.19	0.42	-	****	-	-
N, N-diethyl-S-(4-chloro-2-		-	0.08	0.63	***	0.60	0.64
hydroxybenzyl)thiocarbamate							
N-ethyl-S-(4-chloro-2-hydroxy-		-	0.00	0.50		0.47	0.47
benzyl)thiocarbamate							
N,N-diethyl-S-(4-chloro-3-		**	0.00	0.40	-	0.43	0.52
hydroxybenzyl)thiocarbamate							
N-ethyl-S-(4-chloro-3-hydroxy-			0.00	0.35	-	0.34	0.38
benzyl)thiocarbamate							
N,N-diethyl-S-(4-chlorobenzyl)-		•••	0.00	0.28	-	0.13	0.47
S-monoxythiocarbamate							
4-chlorobenzyl methyl sulfoxide		-	0.00	0.09	***	0.09	0.37
4-chlorobenzyl methyl sulfone		-	0.00	0.33	<u></u>	0.23	0.52

A: hexane only; B: hexane-benzene (1:1); C: chloroform-ether (4:1); D:benzene-ether (1:1); E: benzene saturated with formic acid-ether (3:1), F: benzene-methanol-formic acid (100:20:1)

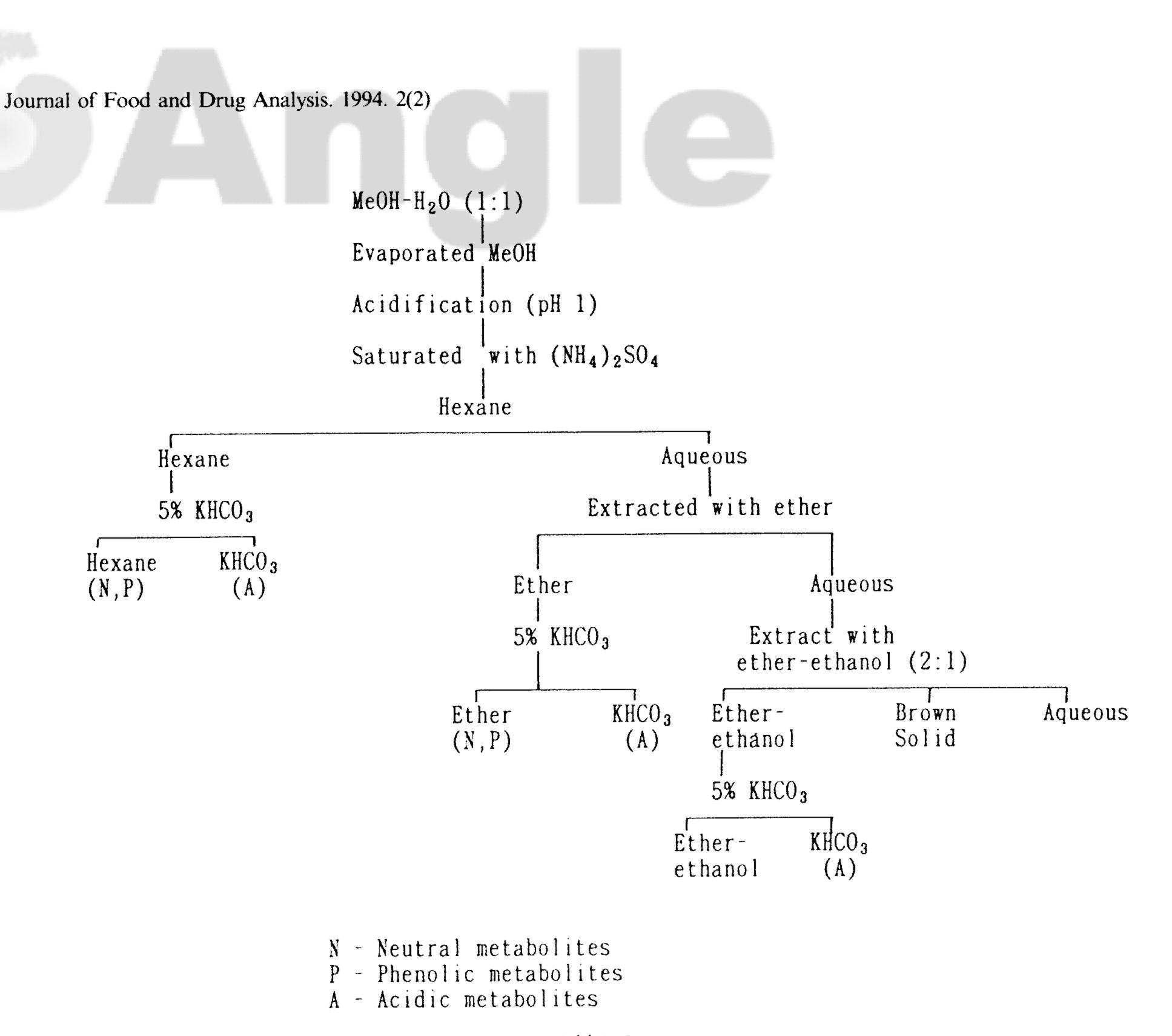


Figure 2. Scheme for fractionation and separation of ¹⁴C from extracts.

and 3% (<0.01 ppm) as 2-hydroxy-4-chlorobenzoic acid. Approximately 5% (<0.01 ppm) of the ¹⁴C in the hexane extract was neutral compound. TLC analysis showed one major spot which corresponded to bis-(4-chlorobenzyl) monodisulfide. This degradation product was probably derived from 4-chlorobenzyl mercaptan by oxidation during the separation process.

2. Ether Soluble ¹⁴C

About 95% (0.38 ppm) of the ¹⁴C in the ether soluble fraction was extractable with 5% potassium bicarbonate solution indicating that most of the ¹⁴C in the ether fraction was also in nature similar to carboxylic acid derivatives. About 30% (0.12 ppm) of the ¹⁴C was isolated and identified as 4-chlorobenzoic acid, and 3% (0.01 ppm) as 2-hydroxy 4-chlorobenzoic acid. There

were two major acidic unknown metabolites designated as "Unknown A" (43%, 0.17 ppm) and "Unknown B" (10%, 0.04 ppm). Unknown A and Unknown B were more polar than 4-chlorobenzoic acid. The Rf values of Unknown A and Unknown B in the benzene (saturated with formic acid)-ether 3:1 (v/v) solvent system were 0.08 and 0.15, respectively, while the Rf value of 4-chlorobenzoic acid was 0.48.

Both Unknown A and Unknown B could be easily methylated with diazomethane in ether solution at 0°C to each form a less polar product. The methylated Unknown A had an Rf value of 0.2 and the methylated Unknown B had an Rf value of 0.46 when developed with chloroform-ether 4:1 (v/v). In this solvent system, both Unknown A and Unknown B (before treatment with diazomethane) stayed at the origin. Unknown A was stable in both 1 N hydro-

chloric acid and 1 N aqueous ethanolic potassium hydroxide solutions at room temperature overnight. However, when Unknown A was heated with 1 N potassium hydroxide in aqueous ethanol solution at 80°C for three hours, it degraded into many radioactive materials which were less polar than the parent compound. The chromatogram was developed with chloroform-ether 4:1 (v/v) for 5 cm followed by hexane-benzene 1:1 (v/v) for 10 cm. The degradation products identified were 20% bis (4chlorobenzyl) disulfide plus 4-chlorobenzyl mercaptan (Rf 0.7) ,12% bis (4-chlorobenzyl) dioxydisulfide (Rf 0.45), 50% bis (4-chlorobenzyl)monoxydisulfide (Rf 0.35), and about 20% stayed at the origin. Mass spectral analysis by electron impact ionization showed no parent (molecular ion) peak in either methylated Unknown A or B, but the base peak of the fragmentation ion at mass 125 (m/e) of the 4chlorobenzyl fragment ion was present. These experimental facts indicate that both unknown metabolites contain 4-chlorobenzylthio and carboxylic acid groups in their molecules. Therefore, it is reasonable to propose that the two metabolites are probably fatty acid derivatives containing the 4-chlorobenzylmercapto group in their molecules.

3. Ether-Ethanol Soluble ¹⁴C

Metabolites in this fraction were mostly conjugated. The ¹⁴C was separated into two fractions with dilute potassium bicarbonate so-

lution. Approximately 80% (0.12 ppm) of the ¹⁴C was extracted with 5% KHCO₃ solution, indicating that most of the ¹⁴C in the ether-ethanol fraction was also acidic in nature. Based on chemical analysis (methylation and hydrolysis) and the Rf values, two major metabolites in this acidic fraction correspond to Unknowns A and B in ether soluble fraction.

The bicarbonate insoluble ¹⁴C (20%) was neutral ether-ethanol soluble ¹⁴C. Studies with acid and α-glucosidase enzyme hydrolysis indicated that about 90% of this ¹⁴C was hydrolyzable. The radioactive products released from this conjugated fraction after enzyme or acid hydrolysis were ether soluble. 4-Chlorobenzoic acid was identified as a major, and 4-chlorobenzyl disulfide was identified as a minor product.

4. Water Soluble ¹⁴C

A small amount of ¹⁴C (4%) remained in the aqueous residue. It was composed of very polar materials, and no further investigation was done.

5. Brown Solid Material ¹⁴C

The brown solid material isolated from the aqueous methanol extract of the straw had high radioactivity. It was obtained as precipitates by saturation of aqueous methanol extract with ammonium sulfate. The concentration of the ¹⁴C in this solid was 17 ppm (554 dpm/mg), about eight times higher than the ¹⁴C concentration in

Table 2. Nature of ¹⁴C in the aqueous methanol extract from straw

Fraction	% of ¹⁴ C	% of 14C	ppm of ¹⁴ C calculated
	in extract	in straw	as benthiocarb
Hexane soluble	7.1	4.4	0.088
Ether soluble	36.1	20.2	0.404
Ether-ethanol soluble	13.2	7.4	0.148
Brown solid material	35.9	20.1	0.402
Aqueous residue	7.1	4.0	0.080
Total	99.4	56.1	1.122

the original straw. This solid material was not soluble in benzene, ether, acetone, dioxane or dimethyl sulfoxide, but it was sparingly soluble in hot alcohol. Elemental and spectral analyses were carried out. Infrared spectra indicated that it contained (A) SiO₂H₂O (3100-3650, 1100 and 800 cm⁻¹); (B) alkyl (2920 and 2860 cm⁻¹); (C) aromatic and unsaturated (1640, 1540, 1460 and 1400 cm⁻¹). Elemental analysis showed that this solid was composed of 35.6% of carbon, 5.4% of hydrogen and 4% of nitrogen. Spectrochemical analysis showed that the solid contained 11.3% of silicon. The analytical results indicated that the ¹⁴C in the brown solid appeared to be natural constituents mainly composed of lignin and silicates derivatives.

Characterizaton of Alkaline Extractable ¹⁴C from the Straw (0.70 ppm) after the Use of Aqueous Methanol

The aqueous alkaline extracts were acidified and fractionated according to the procedure used in the separation of the ¹⁴C in the methanol-water extracts (Figure 2). The fractionation data (Table 3) indicate that approximately 18% (0.13 ppm) of the ¹⁴C in the extract was hexane soluble; about 14% (0.10 ppm) was extractable with ether; about 17% (0.12 ppm) was extractable with ether-ethanol; and less than 1% (< 0.01 ppm) remained in aqueous residue. Approximately 50% (0.35 ppm) of the ¹⁴C in the straw isolated as a brown solid material was a lignin derivative. TLC analysis of the hexane and ether

soluble fractions indicated that about 90% (0.11 ppm) of the ¹⁴C in hexane extract, and about 70% (0.07 ppm) of the ¹⁴C in the ether extract, were isolated as 4-chlorobenzoic acid.

Characterization of ¹⁴C in the Final Residue (0. 18 ppm) of Straw after Successive Extractions with Aqueous Methanol and Alkaline Solutions

Less than 10% of the ¹⁴C (0.18 ppm) remained in the residue after successive extraction with aqueous methanol and alkaline solution. Most of this ¹⁴C could be removed by boiling with 20% sodium bisulfite solution followed by 20% potassium hydroxide solution. The cellulose isolated by this pulping procedure contained about 1% (0.02 ppm) of the total ¹⁴C in the straw, indicating that the majority of the ¹⁴C in the residue was incorporated into the lignin instead of the cellulose fraction.

Nature of ¹⁴C in the Rice Grain (0.3 ppm)

The majority of the ¹⁴C in the grain could not be extracted with organic solvents. Only 3% of the ¹⁴C in the grain was extracted with benzene, 4% with acetone, 8% with methanol and 10 to 12% with aqueous methanol. The nature of the ¹⁴C in the aqueous methanol extract is shown in Table 4. Separation and identification by TLC showed that the ¹⁴C in the hexane and the ether fractions was the same product. It was identified as 4-chlorobenzyl methylsulfone. Therefore, over 60% (0.02 ppm) of the ¹⁴C in

Table 3. Nature of ¹⁴C in the alkaline extract from straw

Fraction	% of 14C	% of MC	ppm of 14C calculated
	in extract	in straw	as benthiocarb
Hexane soluble	18.3	6.4	0.128
Ether soluble	14.3	5.0	0.100
Ether-ethanol soluble	16.6	5.8	0.116
Brown solid material	50.3	17.6	0.354
Aqueous residue	0.6	0.2	0.004
Total	1(X),()	35.0	0.702

the aqueous methanol extract was isolated as 4-chlorobenzyl methyl sulfone. No 4-chlorobenzoic acid was found from grain samples.

Approximately 90% of the ¹⁴C remained in the residue after aqueous methanol extraction. This would be considered as incorporated ¹⁴C. To determine the nature of this ¹⁴C, the grain residue was subjected to drastic alkaline or acid hydrolysis (six hours reflux in 10% KOH or 10% HCl). In the alkaline hydrolysis, the hydrolysate was acidified and extracted with etherethanol 1:1 (v/v). About 40% (0.13 ppm) of the ¹⁴C was organo-extractable and about 50% (0.14 ppm) was organo-unextractable. Only 30% (0.04 ppm) of this organo soluble ¹⁴C was acidic derivatives (potassium bicarbonate extractable), and 70% (0.09 ppm) were neutral or phenolic products (potassium bicarbonate unextractable). In the acid hydrolysis, 40% (0.12 ppm) of the ¹⁴C was organo-extractable and 50% (0.15 ppm) was organo unextractable. In the organo extractable ¹⁴C from the acid hydrolysate, about 20% (0.03 ppm) of the organo soluble ¹⁴C was extracted with dilute potassium bicarbonate and 80% (0.1 ppm) was not. Therefore, approximately 0.03 ppm ¹⁴C acidic materials and 0.1 ppm ¹⁴C were neutral or phenolic materials.

DISCUSSION

Degradation fate and terminal residues of ¹⁴C ring-labeled benthiocarb in mature rice plant are discussed. Samples of mature straw

and grain were used for analysis and characterization. In the straw, the ¹⁴C levels calculated as benthiocarb were summarized as follows: free 4chlorobenzoic acid 0.22 ppm; carboxylic acidic compounds containing the 4-chlorobenzylthio moiety 0.33 ppm; bound ¹⁴C released as 4chlorobenzoic acid after treatment with alkali 0.23 ppm; glucose conjugate of 4-chlorobenzoic acid 0.03 ppm; lignin derivative 1.03 ppm; cellulose fraction 0.02 ppm; very polar materials which remained in aqueous residue 0.08 ppm; 2hydroxy-4-chlorobenzoic acid 0.02 ppm and glucose conjugate of 4-chlorobenzyl mercaptan < 0.01 ppm. The ¹⁴C levels calculated as benthiocarb in the grain were determined to be 4-chlorobenzyl methyl sulfone 0.02 ppm; and bound ¹⁴C (mostly incorporated into lignin derivative) 0.28 ppm. This finding made it possible to suggest tentative metabolic pathways for this herbicide (Figure 3). Benthiocarb probably undergoes hydrolysis of the thiocarbamate ester by hydrolase or esterase to form 4-chlorobenzyl mercaptan as an intermediate. This intermediate is further oxidized by mixed function oxidases and/or dehydrogenase to form 4-chlorobenzoic acid, which is further transformed into a glucose conjugate or incorporated into lignin. The 4-chlorobenzyl group could also be conjugated with amino acids. The glycine conjugate of 4-chlorobenzoic acid which was found as major metabolite in mice and rats was not found in rice⁽¹³⁾.

In the grain, most of the ¹⁴C appeared to be incorporated materials because about 90%

Table 4. Nature of ¹⁴C in aqueous methanol extract from grain

Fraction	% of 14C	% of 14C	ppm of ¹⁴ C calculated	
	in extract	in straw	as benthiocarb	
Hexane soluble	45.7	4.6	0.014	
Ether soluble	25.1	2.5	0.008	
Ether-ethnol soluble	9.1	0.9	0.003	
Brown solid material	15.4	1.5	0.005	
Aqueous residue	4.7	0.5	0.002	
Total	100.0	10.0	0.032	

was unextractable with aqueous methanol. In the aqueous methanol extract, almost all of the ¹⁴C was identified as 4-chlorobenzyl methyl sulfone indicating that biological methylation and oxidation of 4-chlorobenzyl mercaptan occurred in rice grain. 4-chlorobenzyl methyl sulfone has also been found as a major metabolite in rat and mouse tissues⁽¹³⁾.

The uptake, translocation and metabolism of benthiocarb in rice seedings have been studied. Benthiocarb was translocated and possibly degraded easily in the rice seeding. After eight days treatment, only 1% of the parent compound was recovered, and only small amounts of ring hydroxylation of benthiocarb were found. However, in this study no parent compound or other thiocarbamate metabolites were found.

ACKNOWLEDGMENTS

The authors owe great thanks to Mr. Charles Fry for his skillful technical assistance and also to Dr. B. V. Tucker for her good advice

and suggestions. Both were the author's former colleagues at the Ortho Research Center of Chevron Chemical Co., Richmond, California. Their help is deeply appreciated.

REFERENCES

- 1. Casida, J. E., E. C. Kimmel, H. Ohkawa and R. Ohkawa. 1975. Sulfoxidation of thiocarbamate herbicides and metabolism of thiocarbamate sulfoxides in living mice and liver enzyme system. Pesticide Biochemistry and Physiology. 5: 111.
- 2. Hubbell, J. P. and J. E. Casida. 1977. Metabolic rate of the N,N-dialkylcarbamoyl moiety of thiocarbamate herbicides in rats and corn. J. Agric. Food Chem. 25: 404-413.
- 3. Ishikawa, K., I. Okuda and S. Kuwatsuka. 1973. Metabolism of benthiocarb (4-chlorobenzyl N,N-diethylthiolcarbamate) in mice. Agric. Biol. Chem. 37: 165-173.
- 4. Nakamura, Y., K. Ishikawa and S. Kuwatsuka. 1974. Uptake and translocation of benthiocarb herbicide by plants. Agric. Biol.

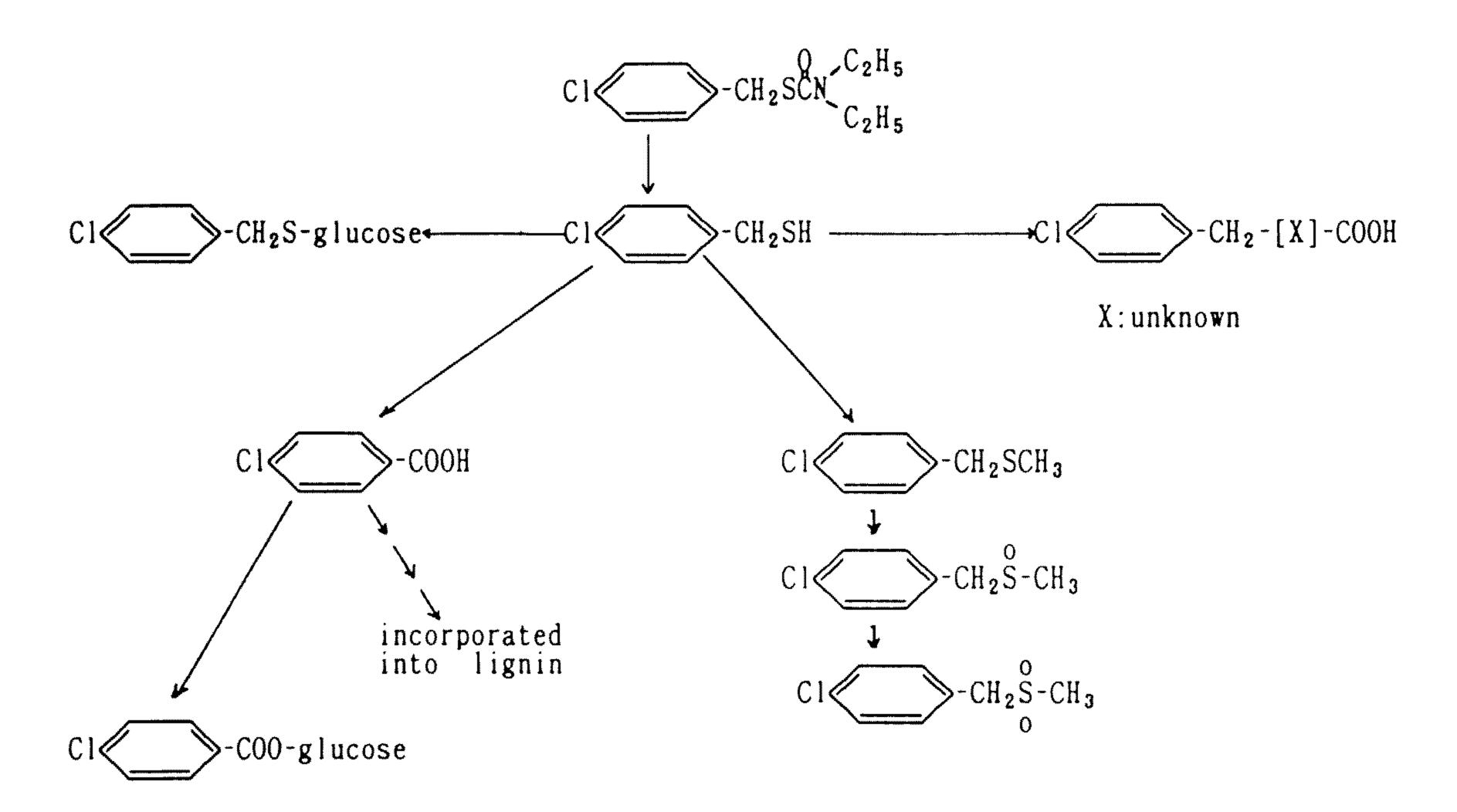


Figure 3. Proposed metabolic pathway of benthiocarb in rice plants.

Chem. 38: 1129-1135.

- 5. Nakamura, Y., K. Ishikawa and S. Kuwatsuka. 1977. Metabolic fate of benthiocarb herbicide in plants. Agric. Biol. Chem. 41: 1613-1620.
- 6. Nakamura, Y., K. Ishikawa and S. Kuwatsuka. 1977. Degradation of benthiocarb in soil as affected by soil conditions. J. Pesticide Sci. 2: 7-16.
- 7. Kuwatsuka, S. and Y. Niki. 1976. Fates and behavior of herbicide in soil environments with special emphasis on the fate of principal paddy herbicides in flooded soils. Review of Plant Protection Research. 9: 143-163.
- 8. Chen, Y. L., H. Fang, L. J. Chen and Y. S. Wang. 1976. Photodecomposition and some behavior of herbicides benthiocarb and DCPA in soils. J. Chinese Agric. Chem. Soc. 14: 59-67.

- 9. Ishikawa, K., Y. Nakamura, Y. Niki and S. Kuwatsuka. 1977. Photodegradaton of benthiocarb herbicide. J. Pesticide Sci. 2: 17-25.
- 10. Cheng, H. M. 1990. Identification of benthiocarb degradation products in chlorinated water. J. Chinese Agric. Chem. Soc. 28: 77-85.
- 11. Cheng, H. M. and D. F. Hwang. 1990. Dissipation of benthiocarb herbicide in deionized water containing chlorine. J. Chinese Agric. Chem. Soc. 28: 296-300.
- 12. Browing, B. L. 1967. Methods of Wood Chemistry, Vol. 2, Chapters 19, 32. New York, John Wiley & Sons Inc.
- 13. Cheng, H. M. and D. F. Hwang. 1993. Metabolism of ¹⁴C-ring labeled benthiocarb in mice and rats. J. Chinese Biochem. Soc. 22: 27-35

除草劑農藥殺丹(Benthiocarb)在水稻植物中最終產物之分離鑑定

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摘 要

除草劑農藥benthiocarb是植物萌前之選擇性農藥,近二十年來被廣泛使用於水稻田以除雜草,爲知benthiocarb在水稻中之命運,乃以5磅/英畝之量撤佈¹⁴C-benthiocarb於水槽缸,缸內種植水稻成熟收割後,探集稻草和稻米來分析¹⁴C量,得知稻草和稻米之¹⁴C全量(以benthiocarb計算)各爲2.0和3.0 ppm。其中均未檢出含有母體benthiocarb和具有thiocarbamate構造體之代謝產物(<

0.01 ppm)存在。稻草中之主要benthiocarb產物爲4-chlorobenzoic acid之自由型和結合型的代謝產物 (0.25 ppm),其次爲含有carboxylic acid和4-chlorobenzylthio構造體之代謝產物(0.33 ppm)。Benthiocarb在稻米中之主要產物爲木質素(lignin)結合物 (0.28 ppm)和4-chlorobenzyl methyl sulfone (0.02 ppm)。

Accepted for Publication: Mar. 4, 1994